



Protein adsorption to lipid membranes through metal ion chelation studied by X-ray and neutron reflectivity and GIXD

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Outline

I. Introduction - motivation, description of lipid/protein system

II. Results

a. Protein conformation / orientation (NR, XR)

-final state

-evolution of layer structure with time

b. Evidence for two stages (GIXD, NR, XR)

-stage 1: reversible

-stage 2: irreversible

III. Summary



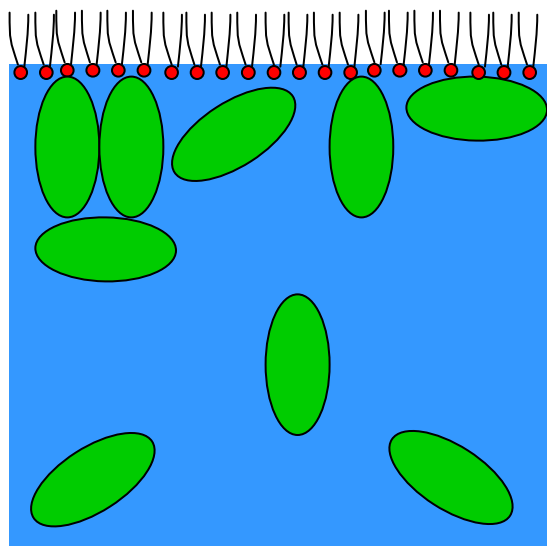
Introduction

Motivation for studying interaction of proteins with lipid membranes (membrane-associated, not integral membrane proteins)

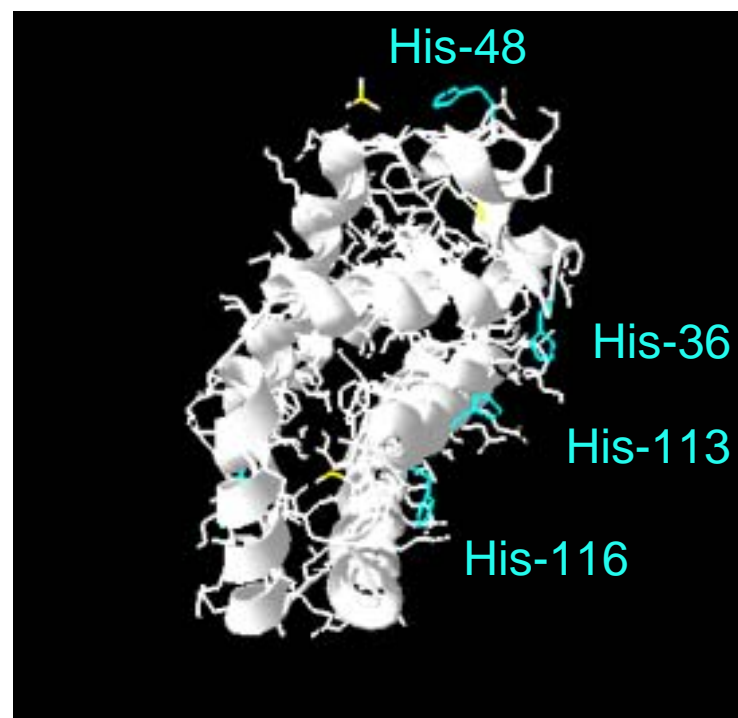
- a). biochemical processes: protein binding and conformational changes regulate ions channels, play important roles in cellular communication, immune response, etc.
- b). nanoscience: control/direct the formation and growth of supramolecular structures (motor protein highways, protein complexes)
- c). mechanisms of toxin assault on cell membranes
- d). biosensors - binding modes determine chemical signals, dictate sensor response, orientation of antibodies

Metal-ion coordination with histidines

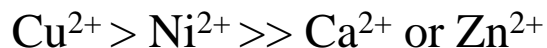
Langmuir monolayers of metal-chelating lipids



myoglobin (horse heart)



strong interaction between histidines and divalent metal ions:





Metal-ion coordination with histidines

-Used for protein separation and purification:

recombinant proteins with “His” tags

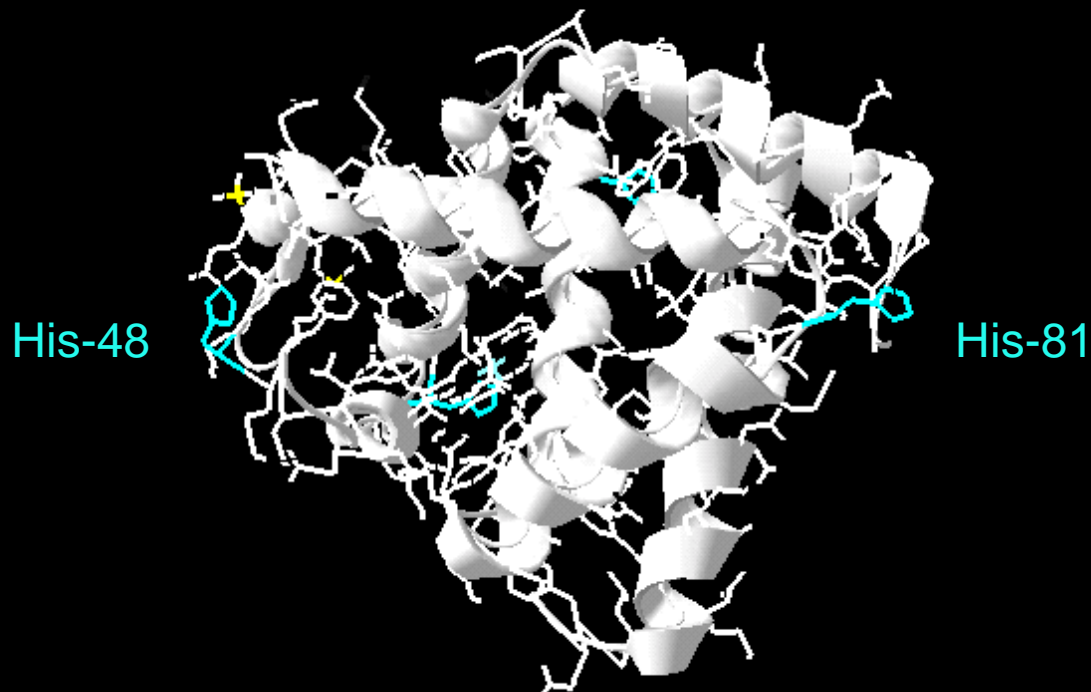
naturally occurring proteins with surface-exposed histidines
can act as contaminants on chromatographic columns of this type

in some cases the goal is to purify naturally occurring proteins
with surface-exposed histidines

-A general method for creating biofunctionalized surfaces

Fundamental understanding of adsorption process needed to:
control orientation, tune energetics (selectivity), avoid denaturation

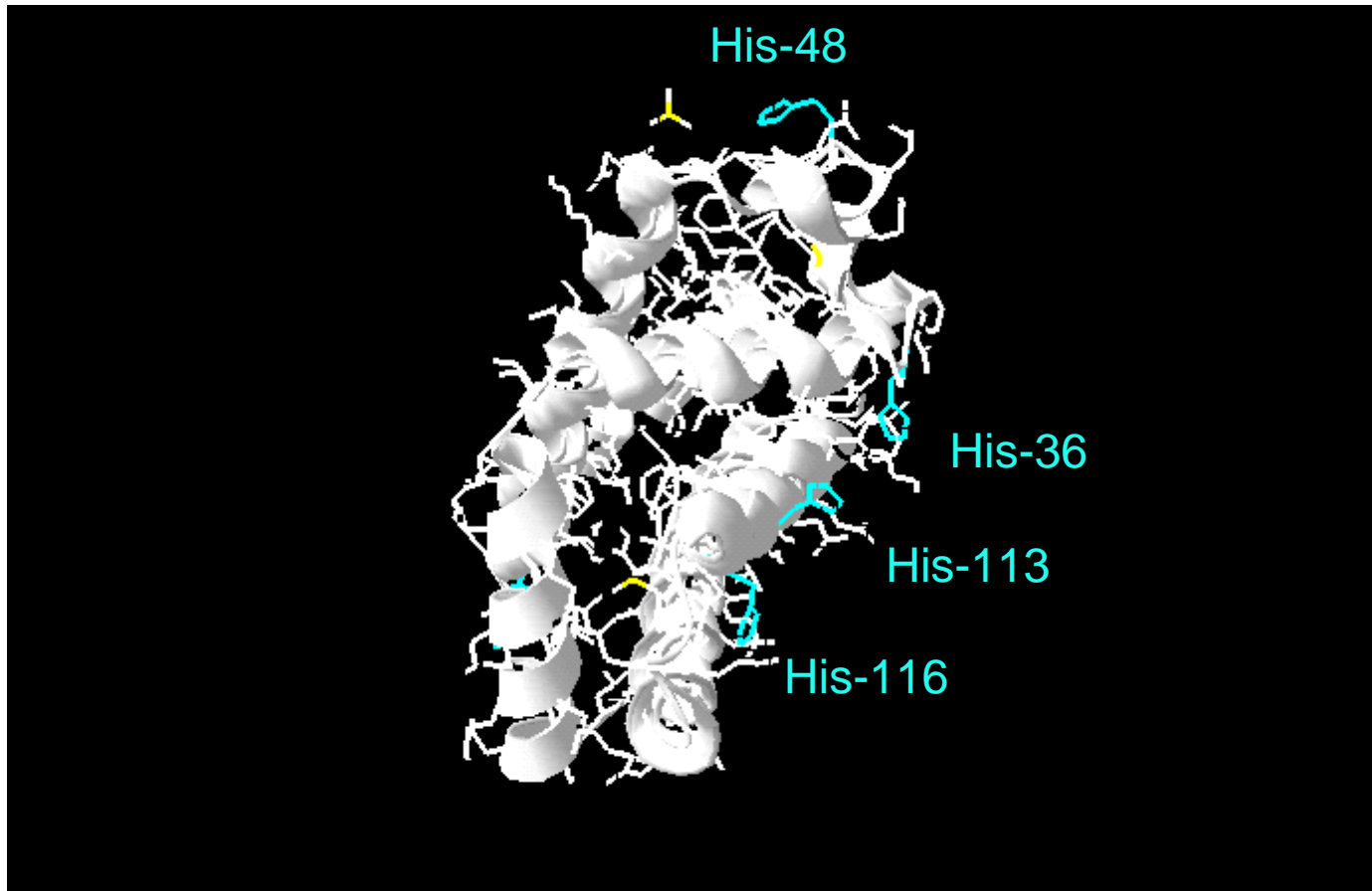
Structure of myoglobin



Dimensions [\AA]:
44 x 44 x 20

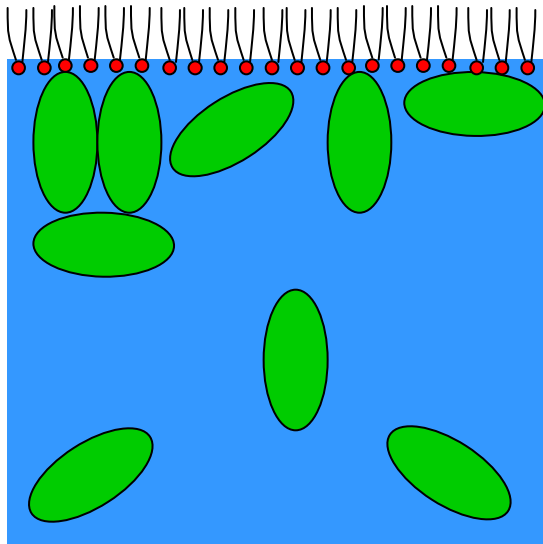
orientation of adsorbed protein depends upon the distribution of histidines

Structure of myoglobin

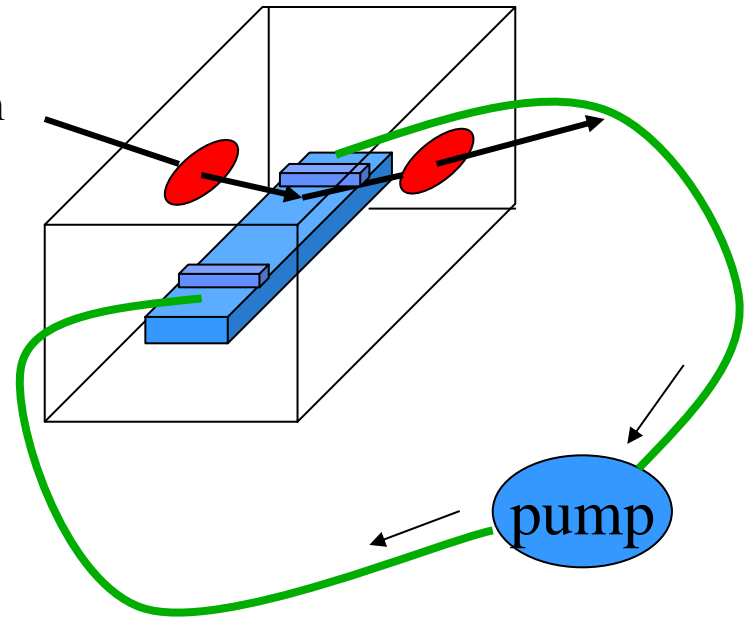


orientation of adsorbed protein depends upon the distribution of histidines

Experimental set-up

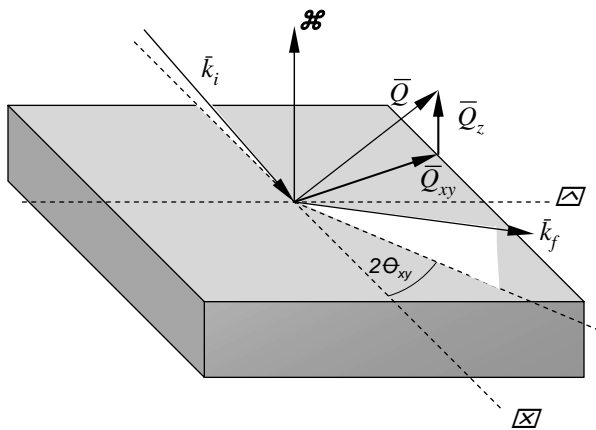


neutron or
X-ray beam

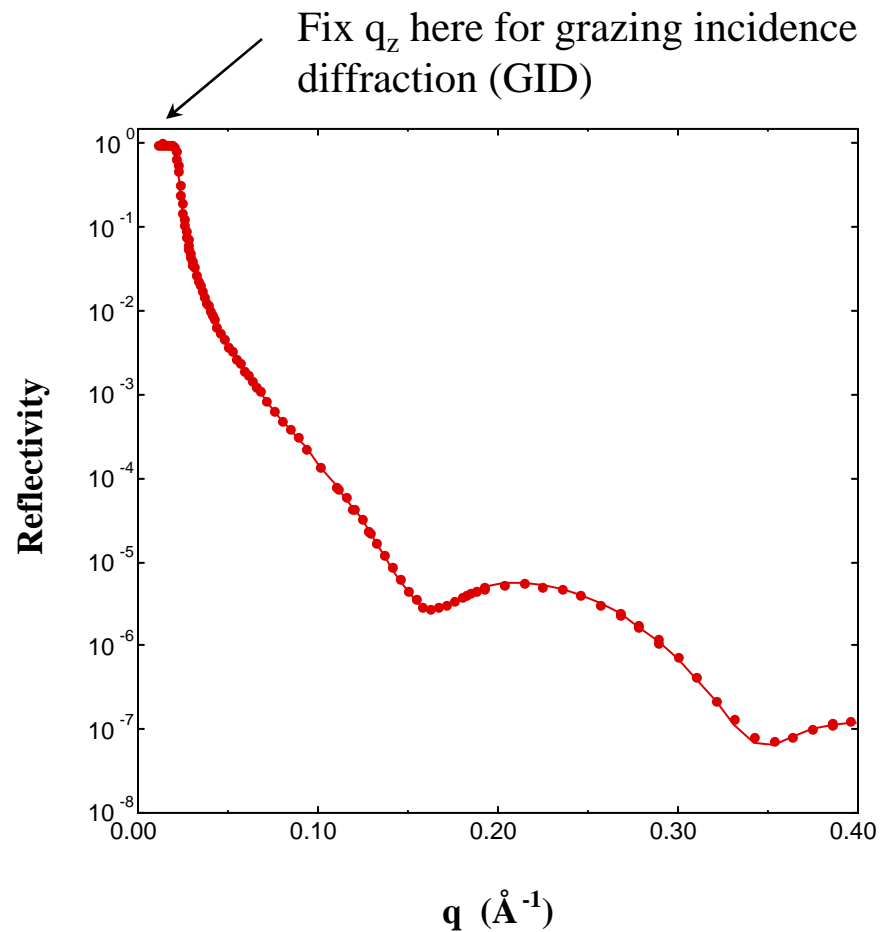


circulate metal ions and myoglobin into the subphase
underneath the lipid layer

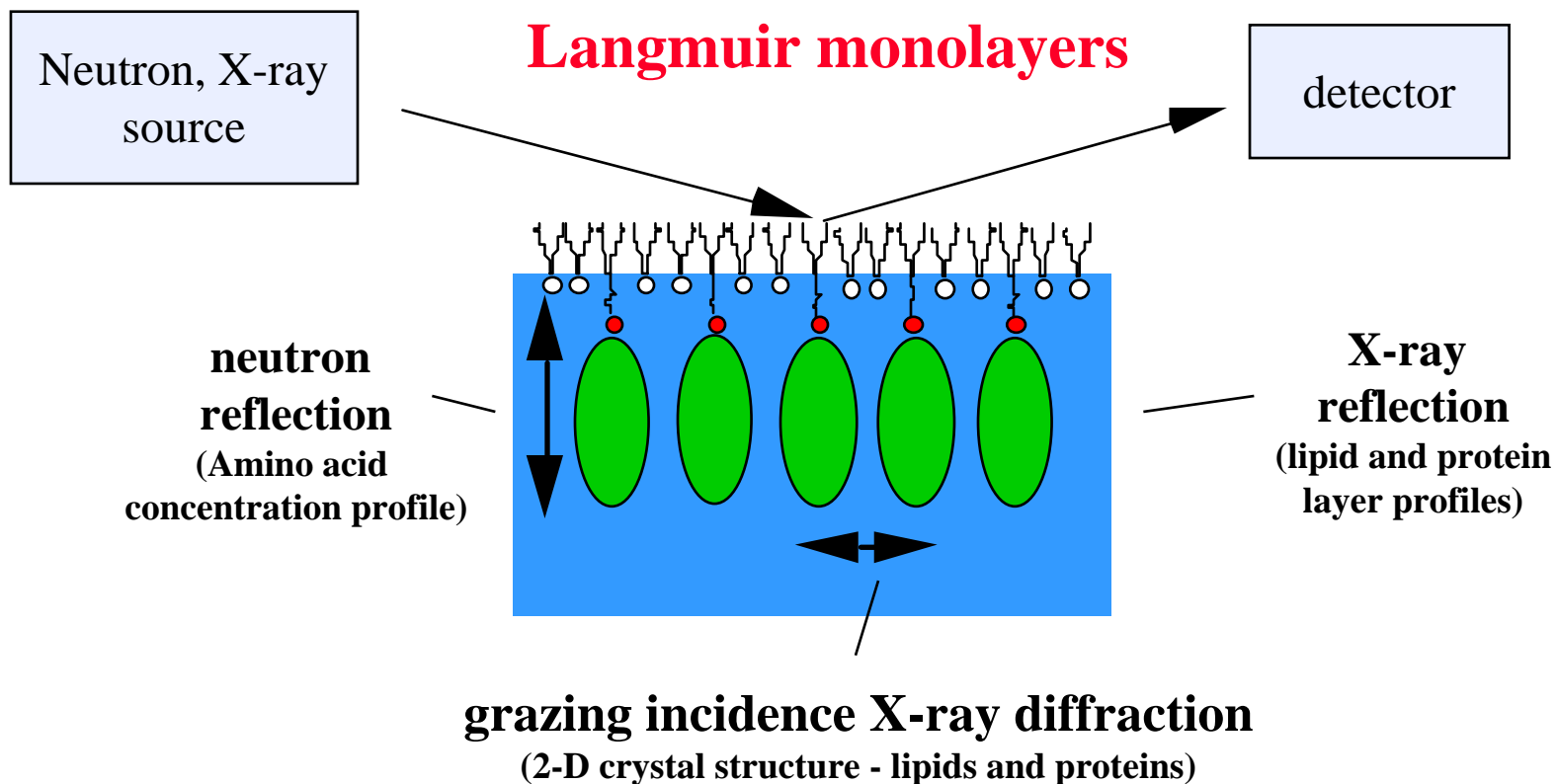
X-ray and neutron grazing incidence scattering techniques



neutrons: reflection only
X-rays: reflection and diffraction



X-ray and neutron grazing incidence scattering techniques





Results

a. Protein conformation / orientation (NR, XR)

- final state

- evolution of layer structure with time

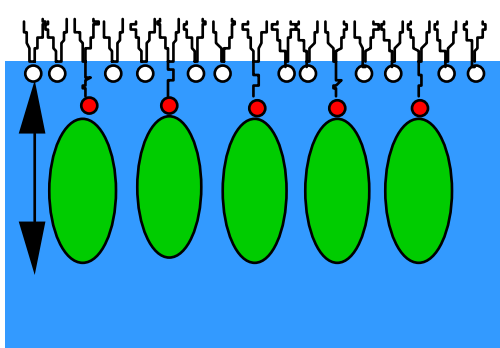
→ b. Evidence for two stages (GIXD, NR, XR)

- stage 1: reversible

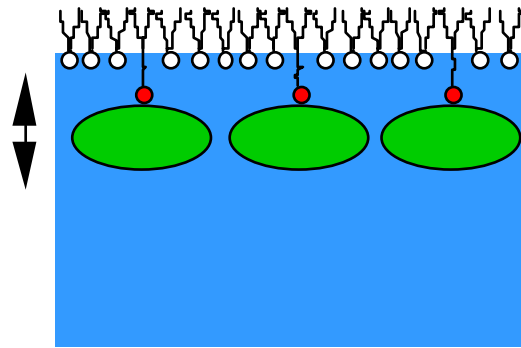
- stage 2: irreversible

Results: A. protein conformation

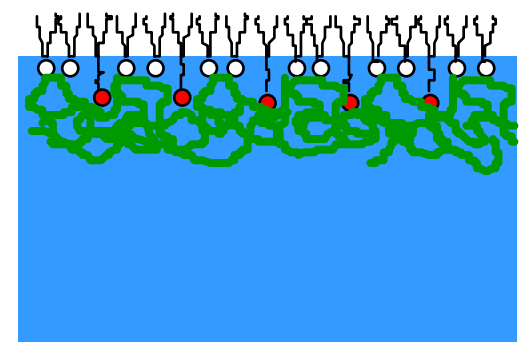
neutron (and X-ray) reflection
probes amino acid segment profile



end-on



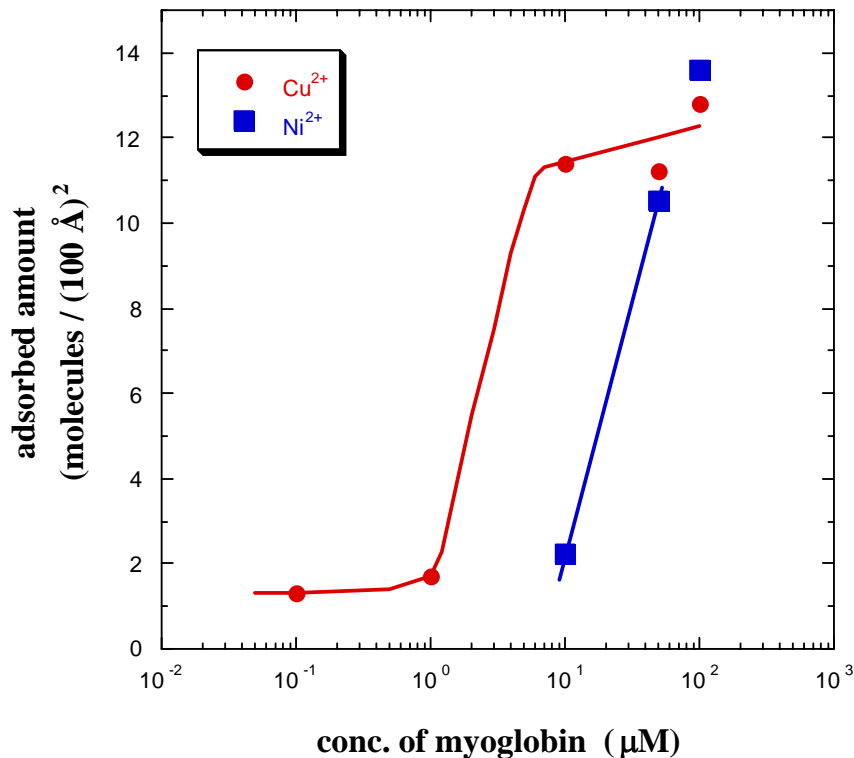
side-on



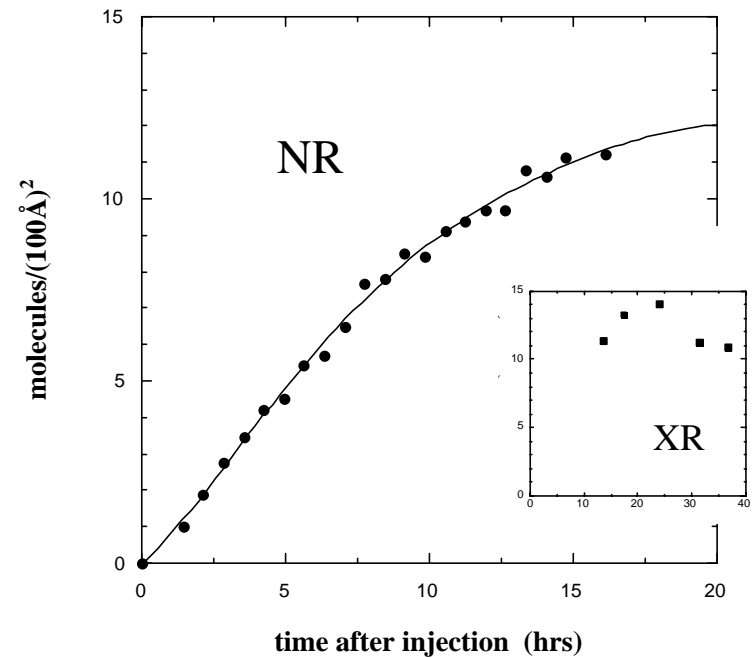
denaturation

Results - surface pressure

“final” state



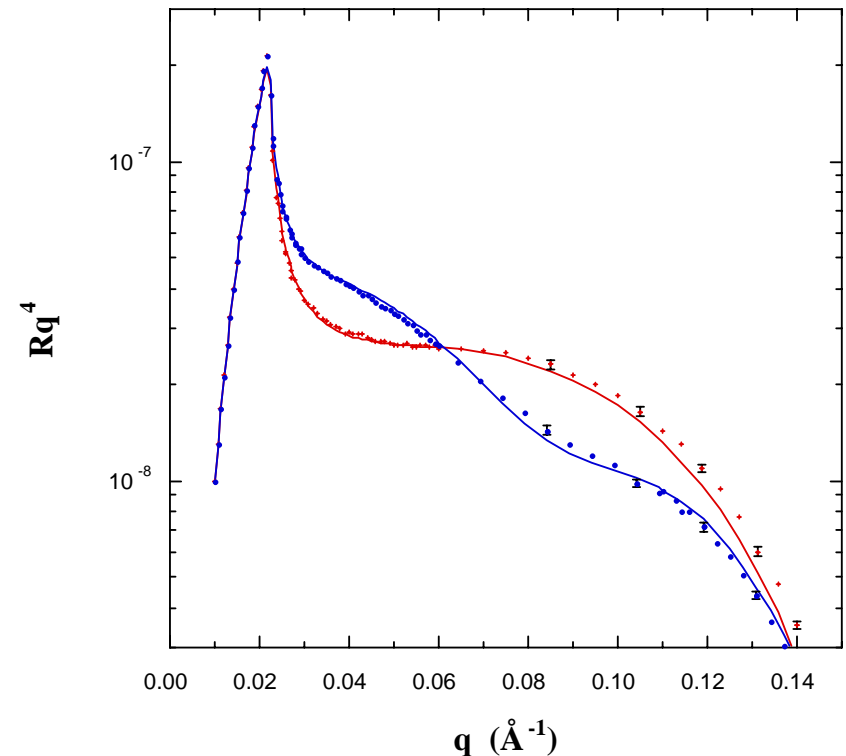
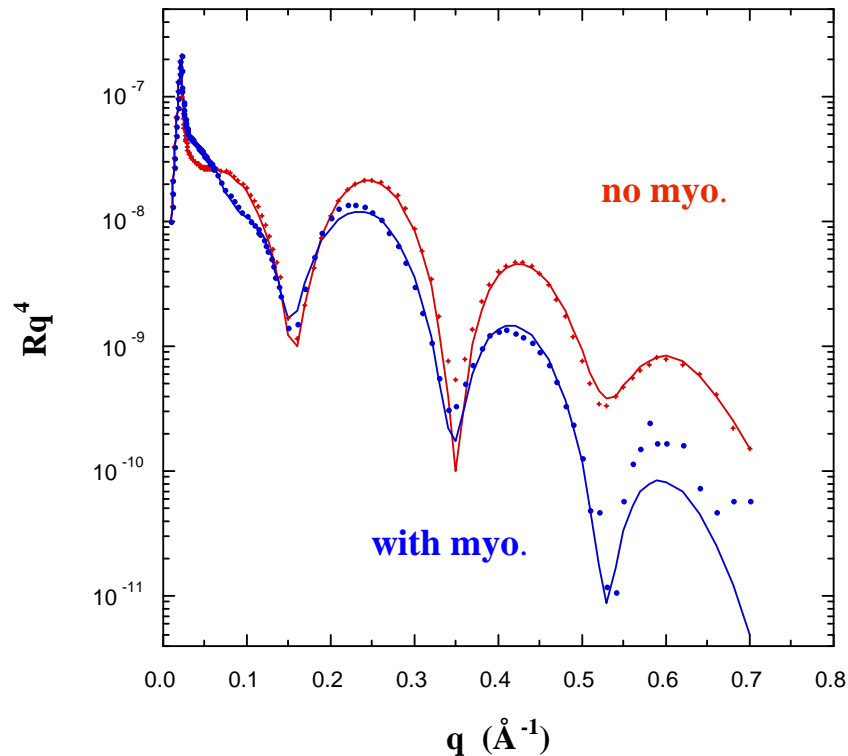
time dependence



Slow kinetics allow study of evolution of protein layer during the adsorption process!

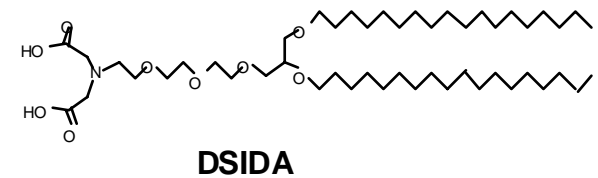
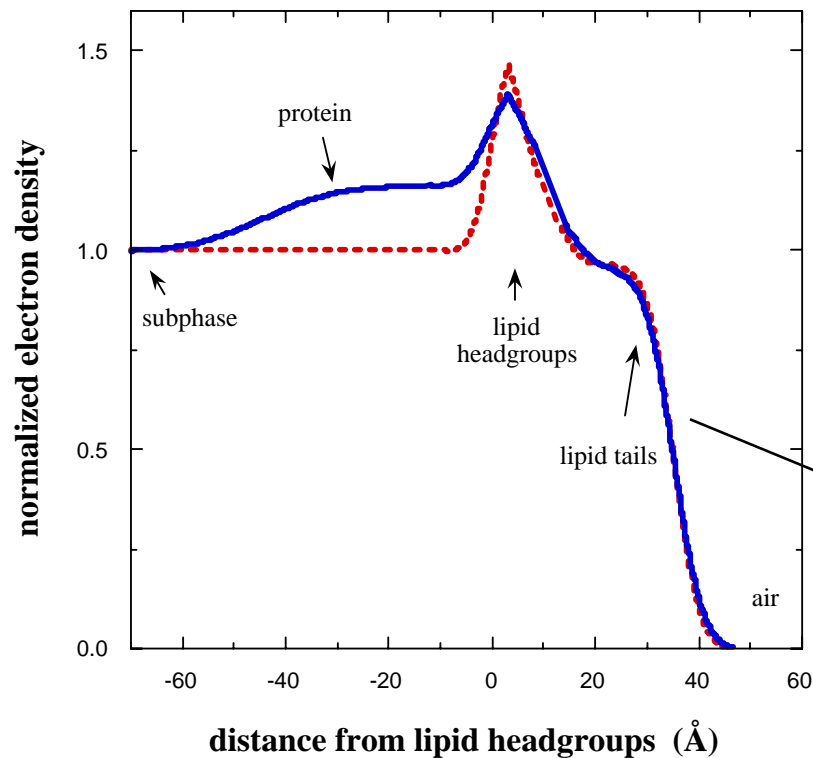
Results - X-ray reflection

chelated Cu^{2+} ions, 10 μM myo.



In-house X-ray source is sufficient to get the dimension of the protein layer in the final state!

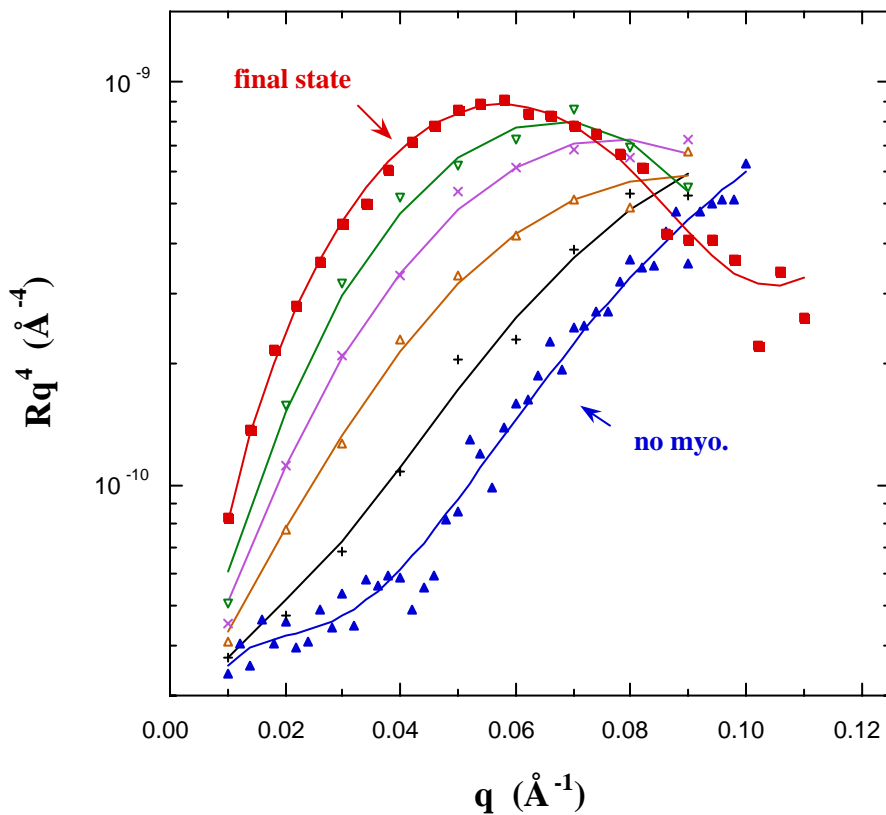
Results - X-ray reflection



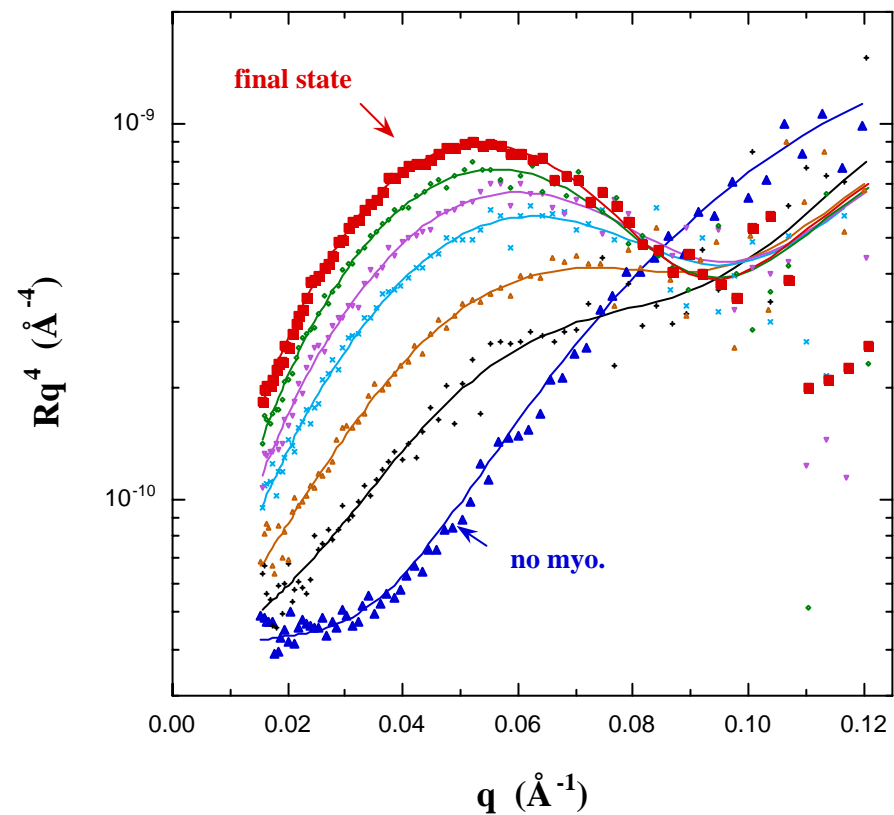
tail layer constrained
by area/molecule
and # of electrons

Neutron reflection, H₂O (time dependence)

Cu²⁺ ions



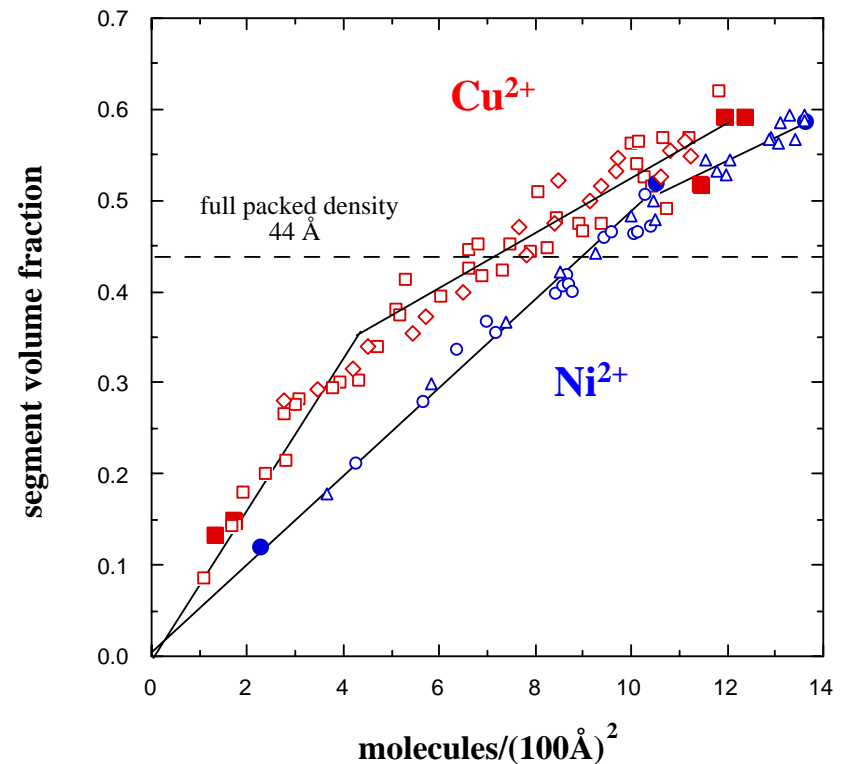
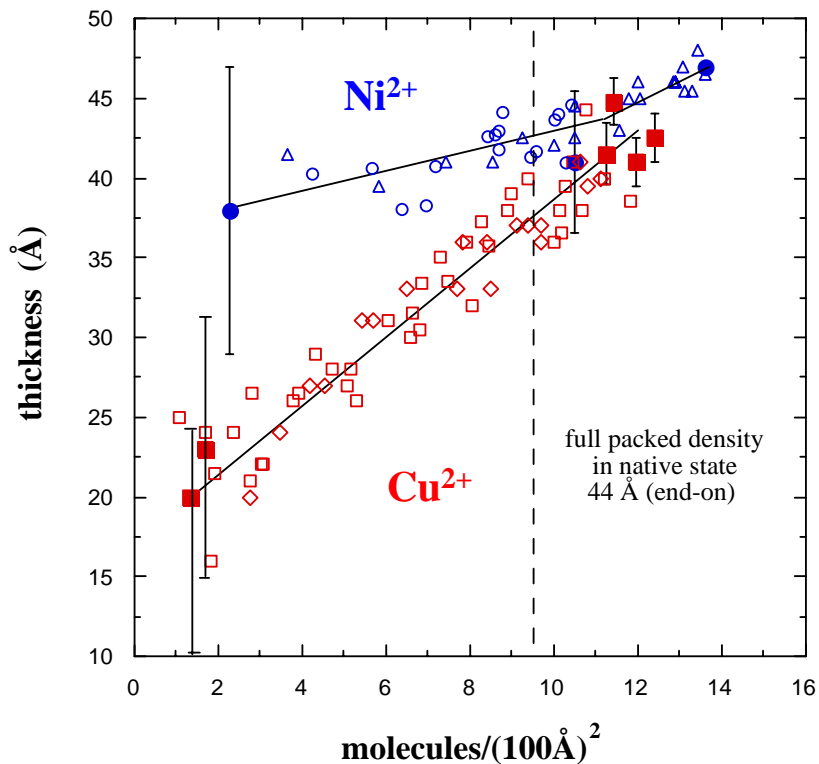
Ni²⁺ ions



Each curve obtained in roughly 40 minutes

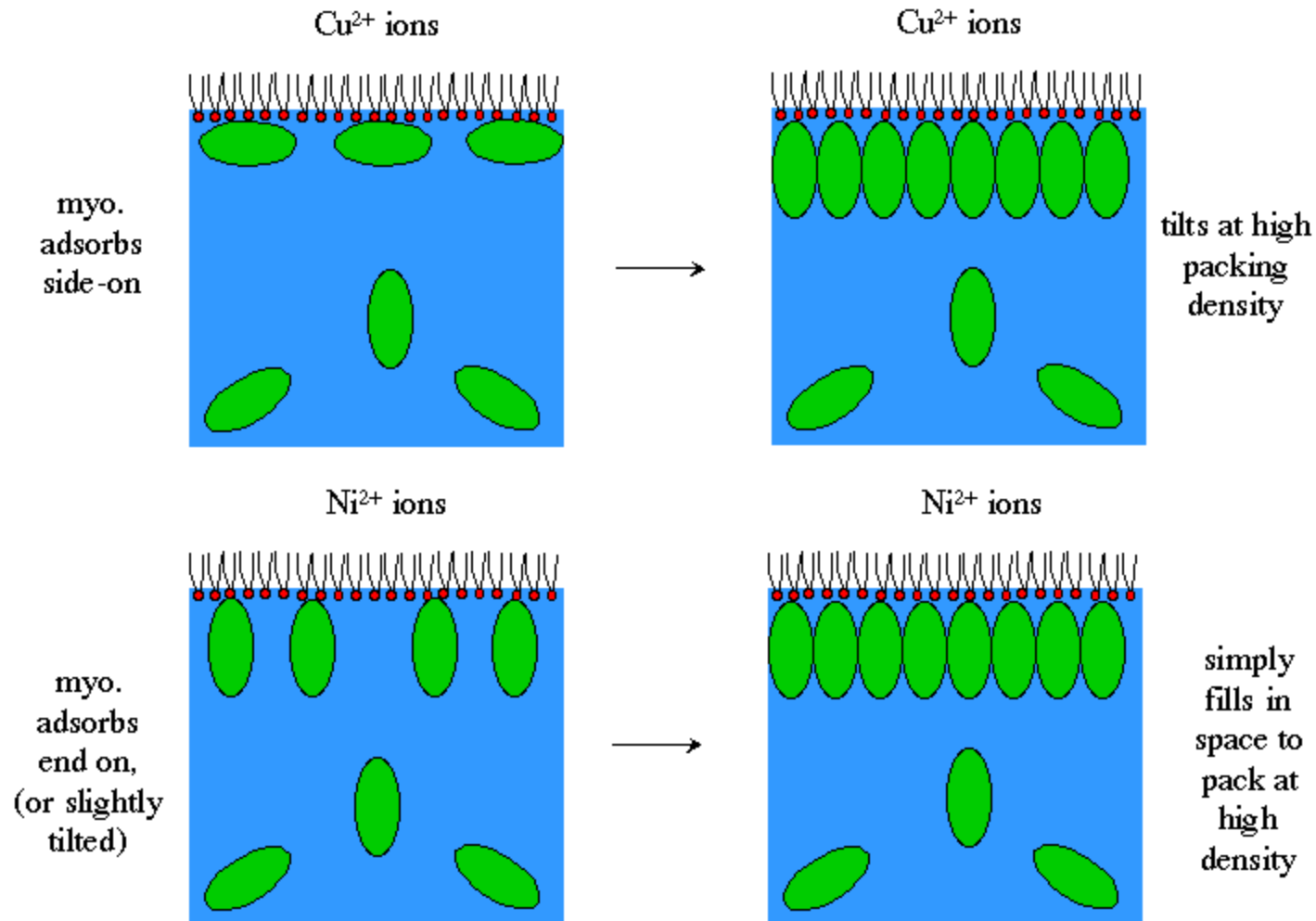
Summary

myoglobin dimensions [\AA]: $44 \times 44 \times 25$

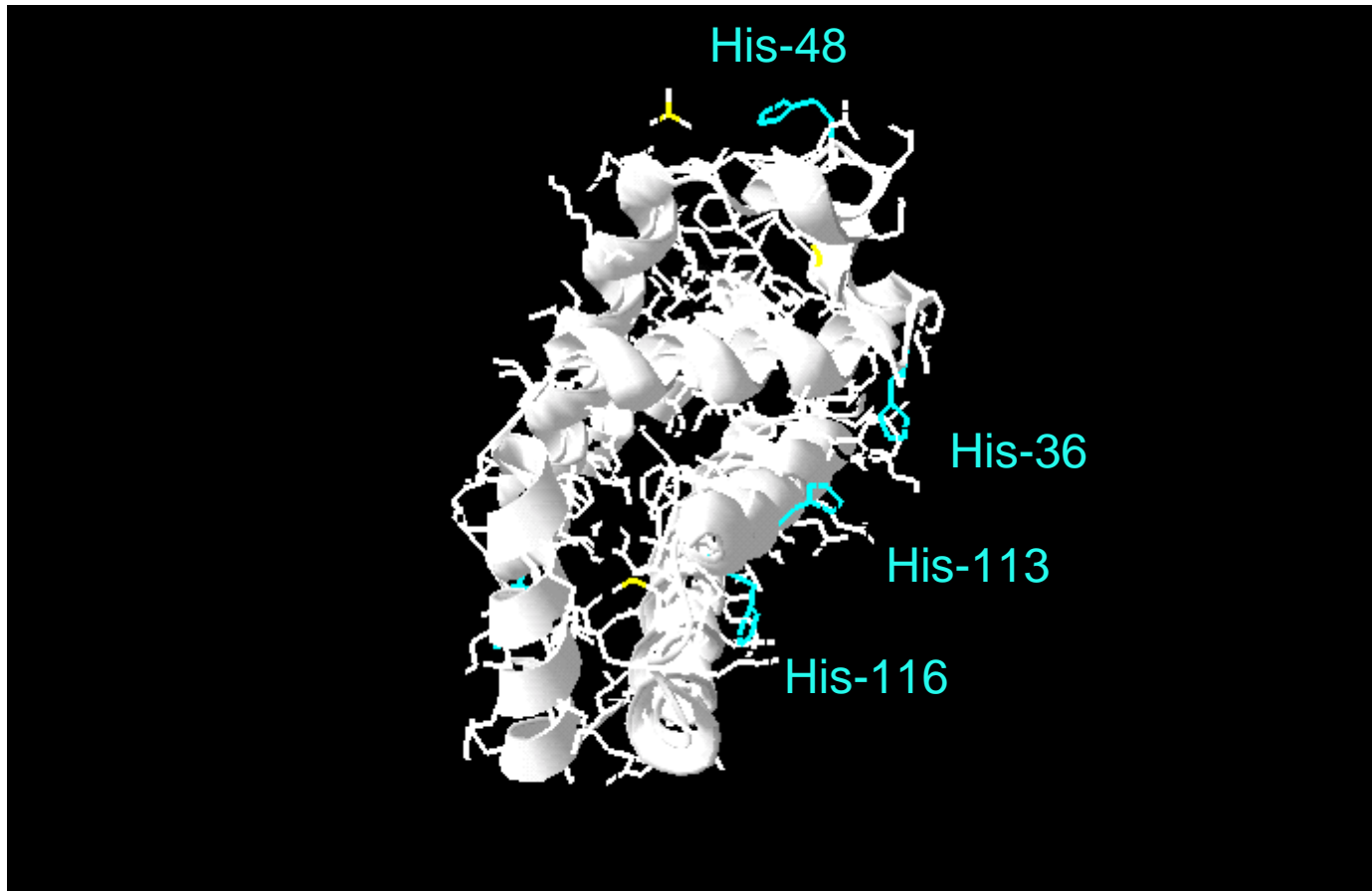


Isolated chains adsorb in a much thinner layer with Cu^{2+} than with Ni^{2+}

Possible interpretation



Possible interpretation

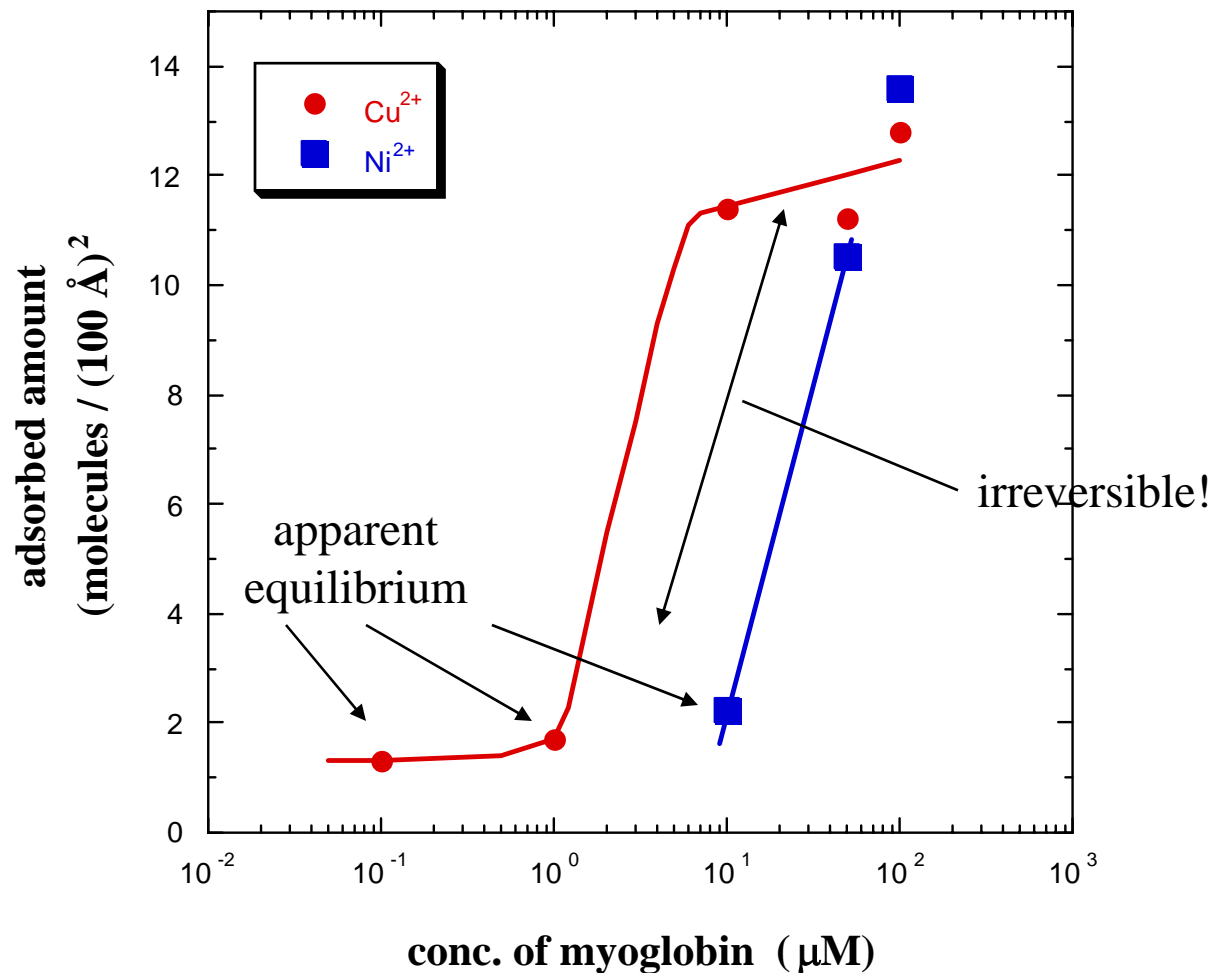


Binding by multiple histidines in case of Cu^{2+} , but His-48 or His-81 for Ni^{2+}

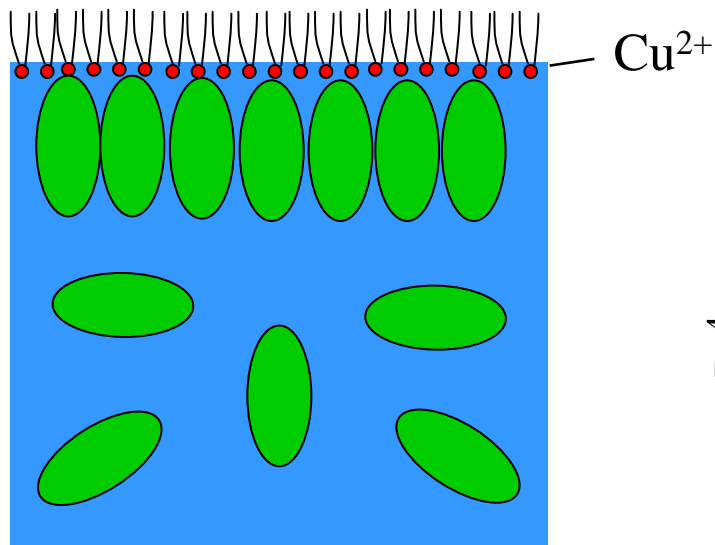
Results: B. Evidence for two stages

Evidence #1.

“final” state

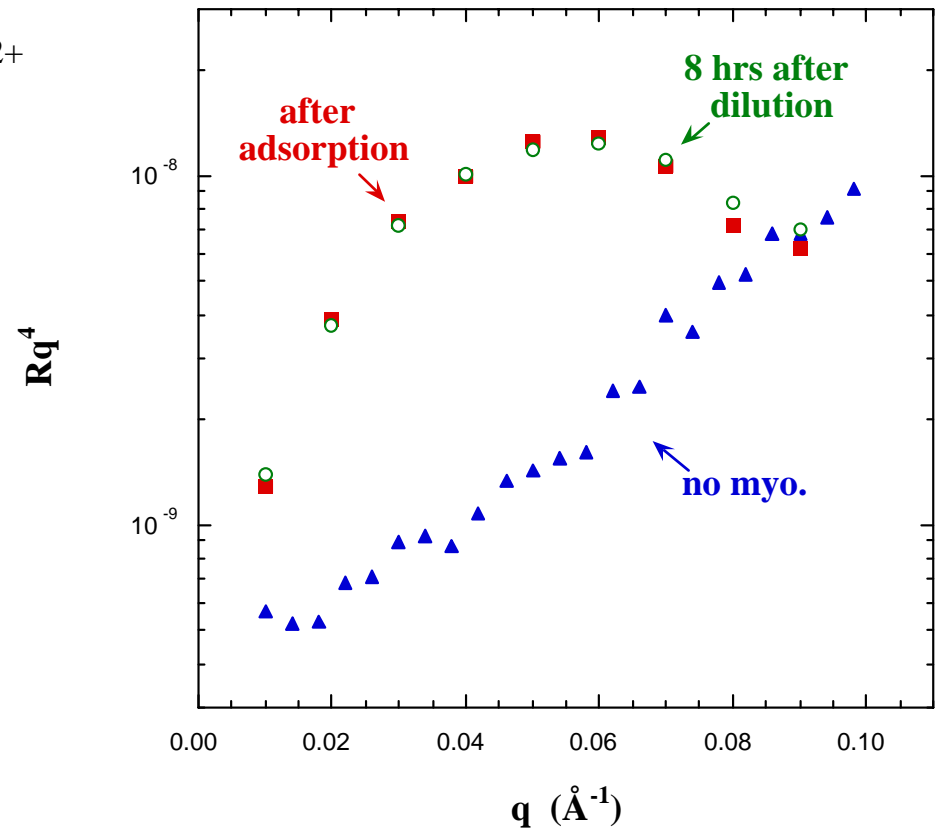


Subphase exchange shows irreversibility



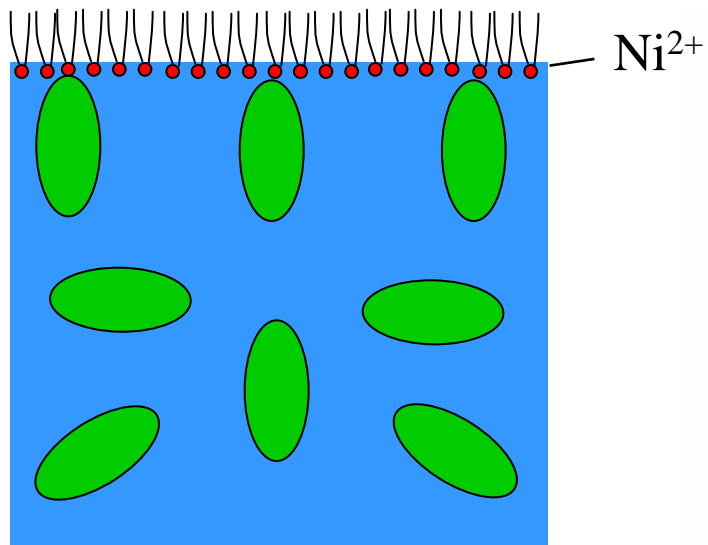
diluted from
50 μM to 1.4 μM

50 μM myo., $\text{Cu}(\text{II})$



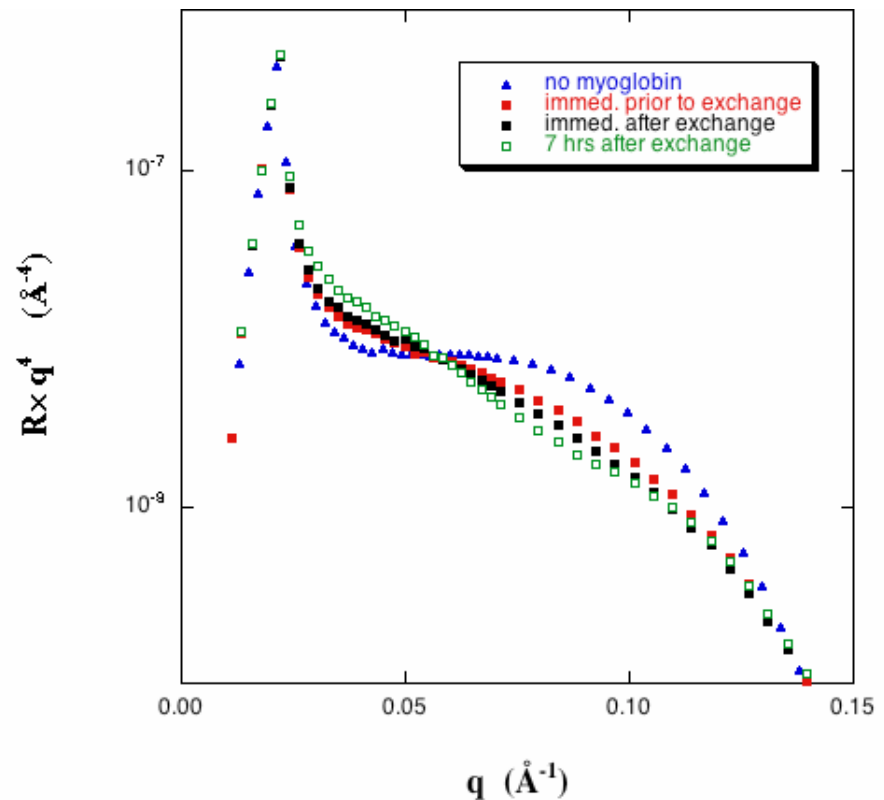
Irreversible in fully packed state!

Subphase exchange shows irreversibility



diluted from
60 μM to 10 μM

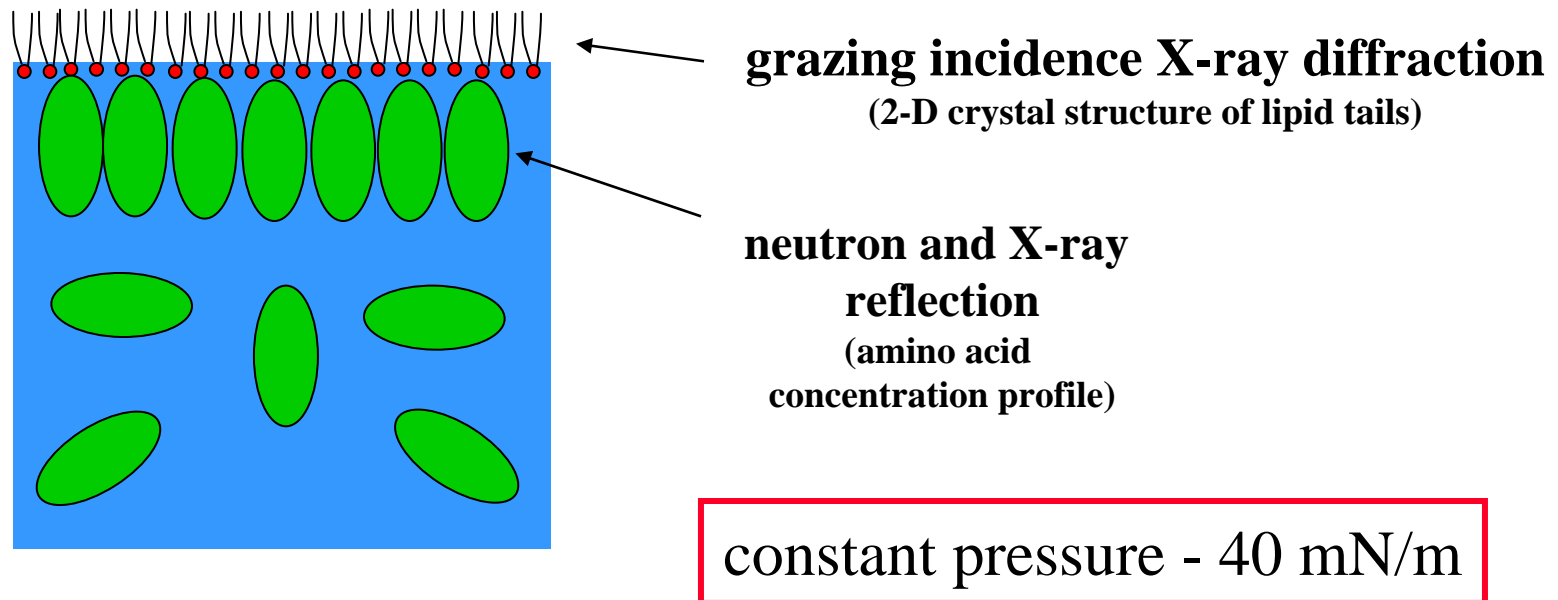
60 μM myo, Ni^{2+}



Irreversible even at moderate coverage with Ni^{2+}

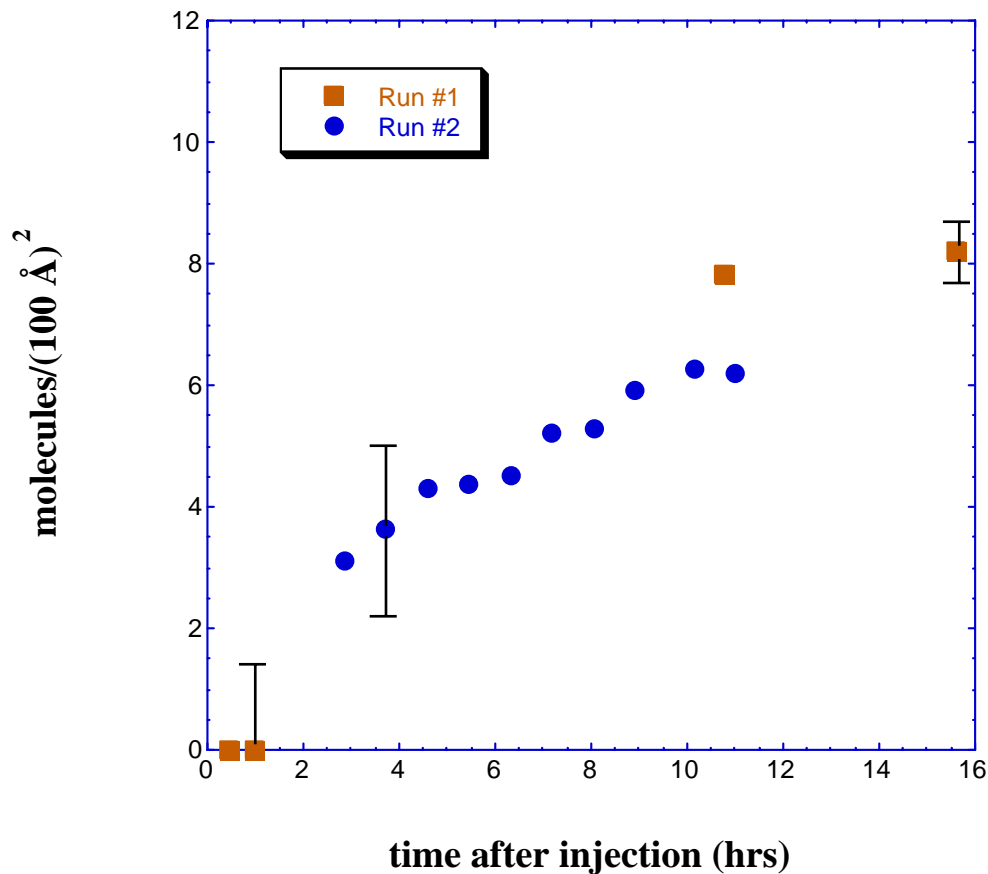
Results: B. Evidence for two stages

Evidence #2. Different time scales for disruption of lipid packing structure and accumulation of adsorbed protein



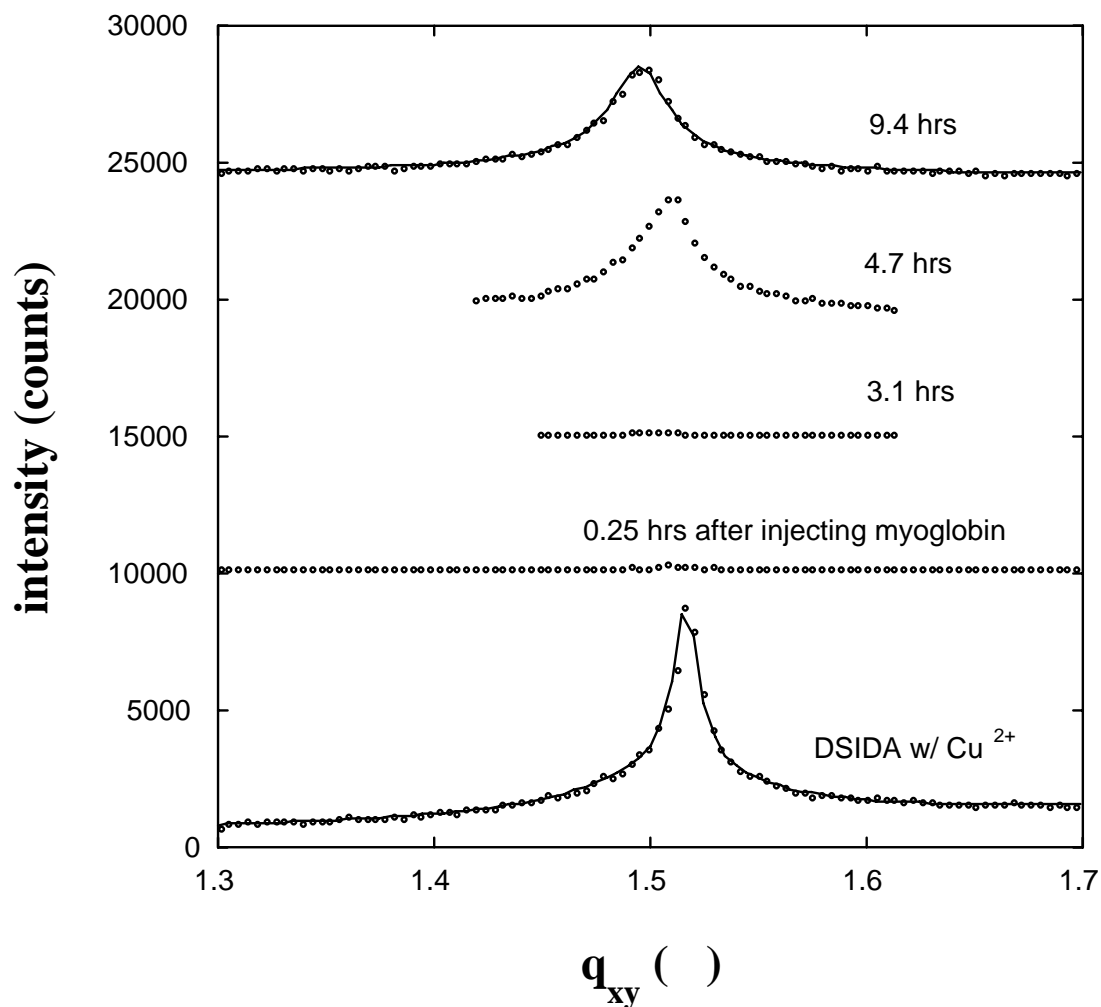
Constant pressure

$\Pi = 40 \text{ mN/m}$, $10 \text{ } \mu\text{M}$ myo.



2 hrs after injection
very little protein
has adsorbed!

Cu^{2+} , 40 mN/m - Bragg Peak

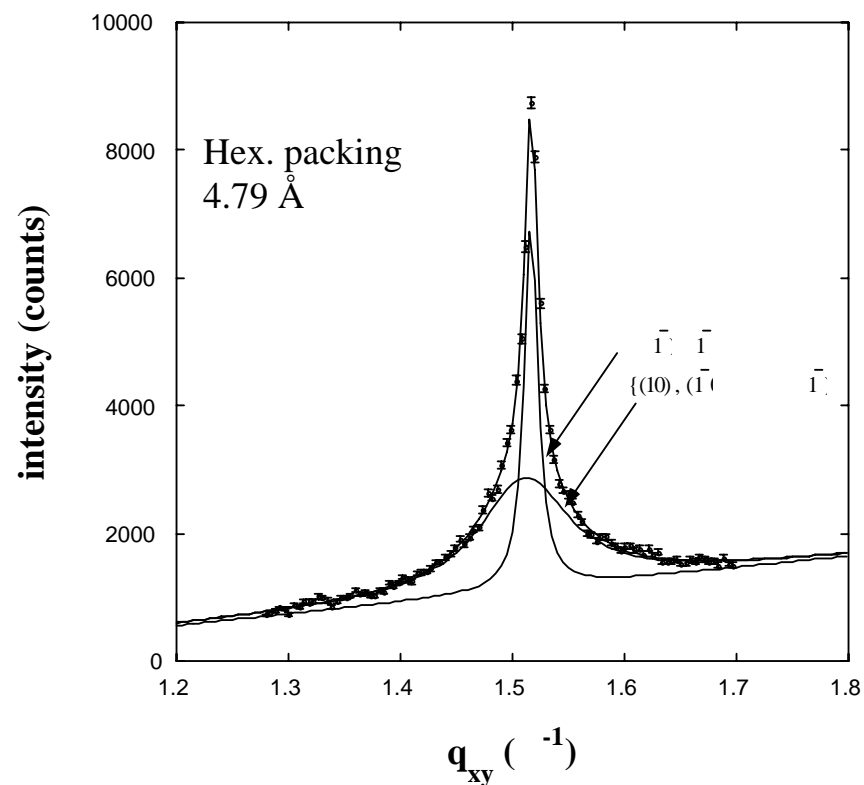


At 0.25 hr after injection, crystallinity is no longer detected in the film!

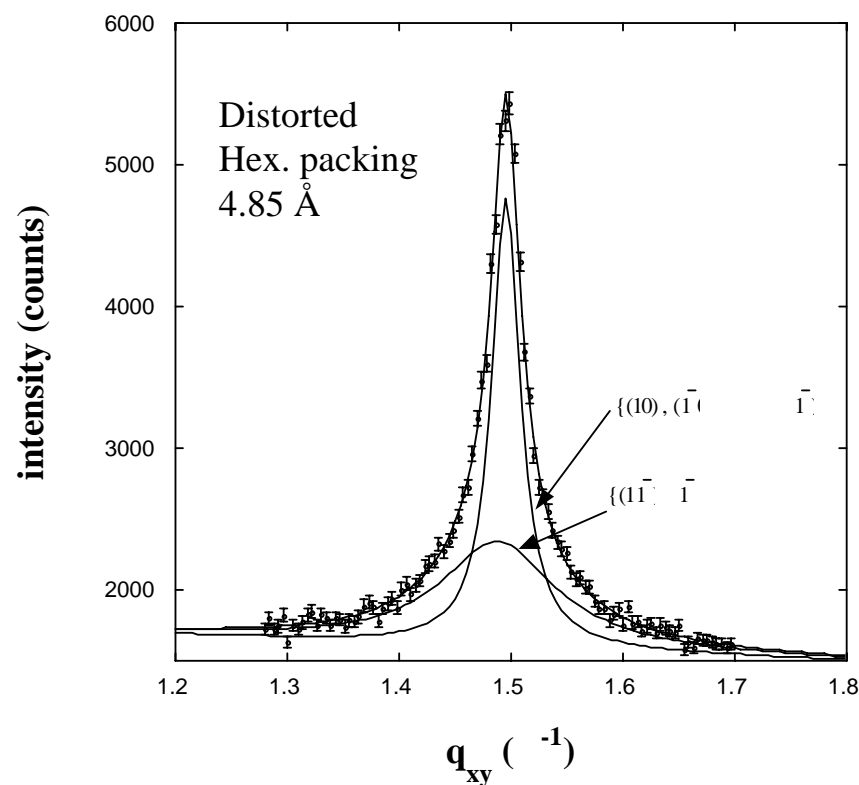
Crystalline packing returns at ~ 5 hrs after injection

Constant pressure - Bragg Peak

Before adding myoglobin

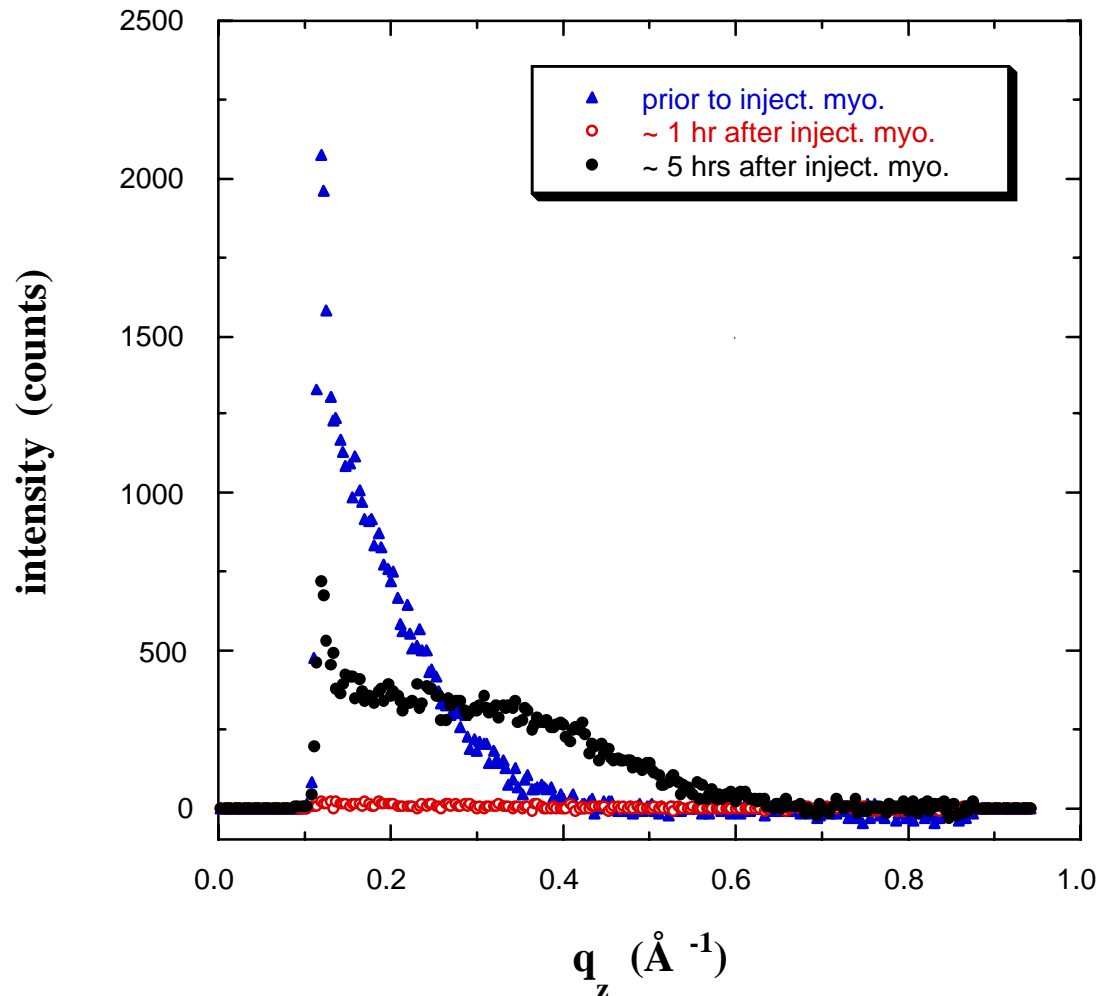


9.4 hrs after adding myoglobin



Diffraction peak returns: broader and at lower q

Constant pressure - Bragg Rod



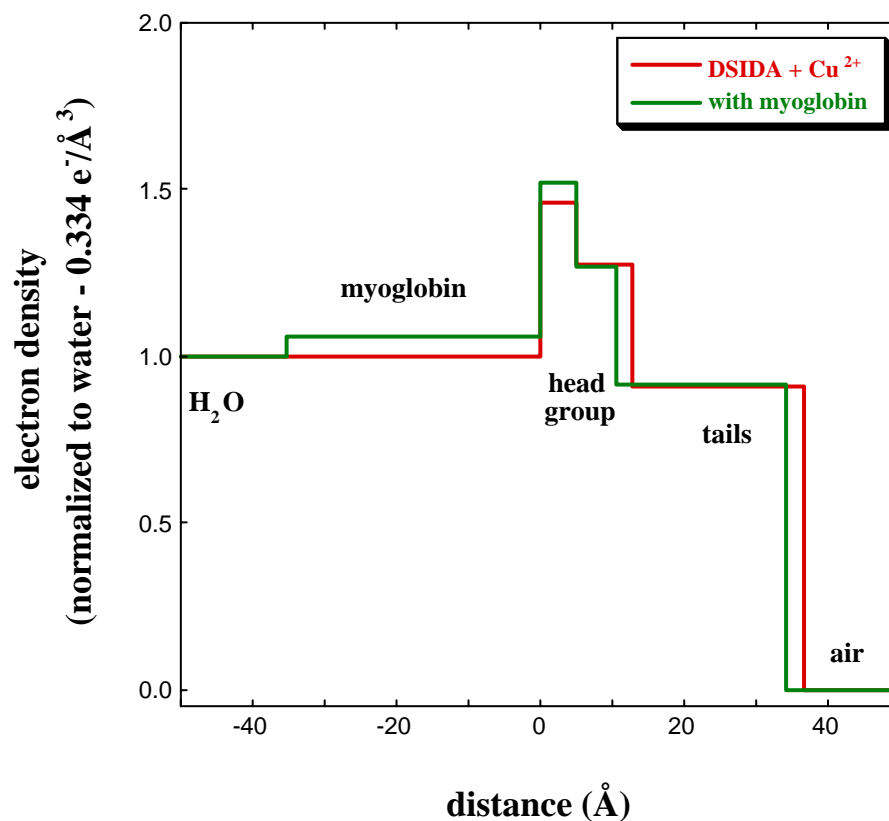
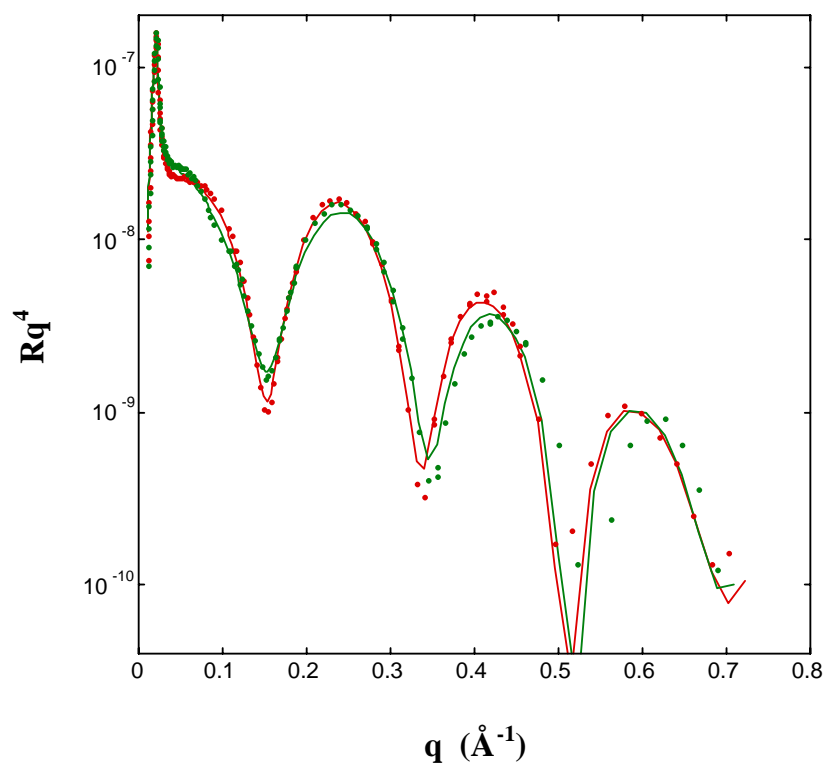
Prior to injection, tails are vertical, crystalline packing

At 1 hr after injection, no crystallinity is detected in the film!

Crystalline packing returns at ~ 5 hrs after injection - tails are tilted

Constant pressure

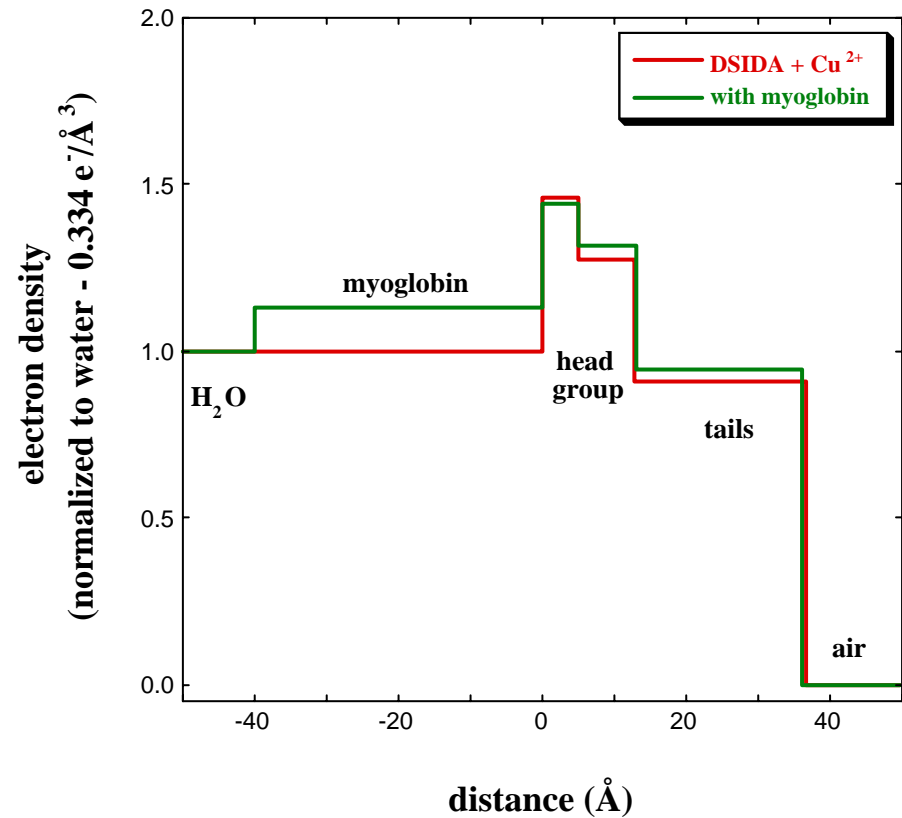
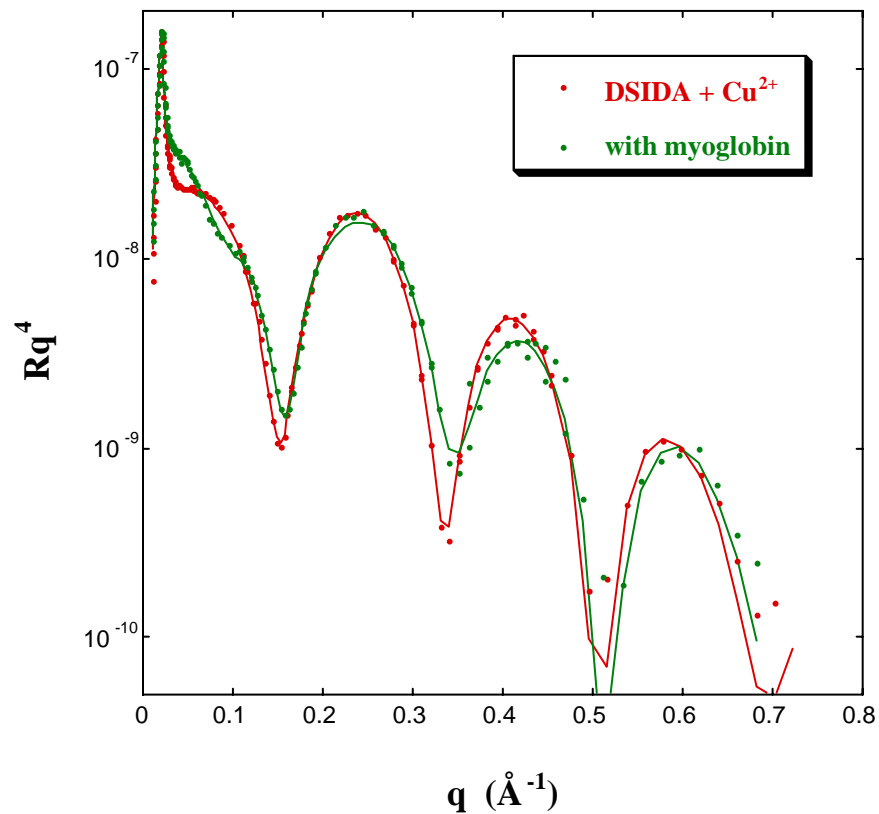
0.9 hrs after injecting myoglobin



very little adsorbed protein: thick. and vol. fract. uncertain

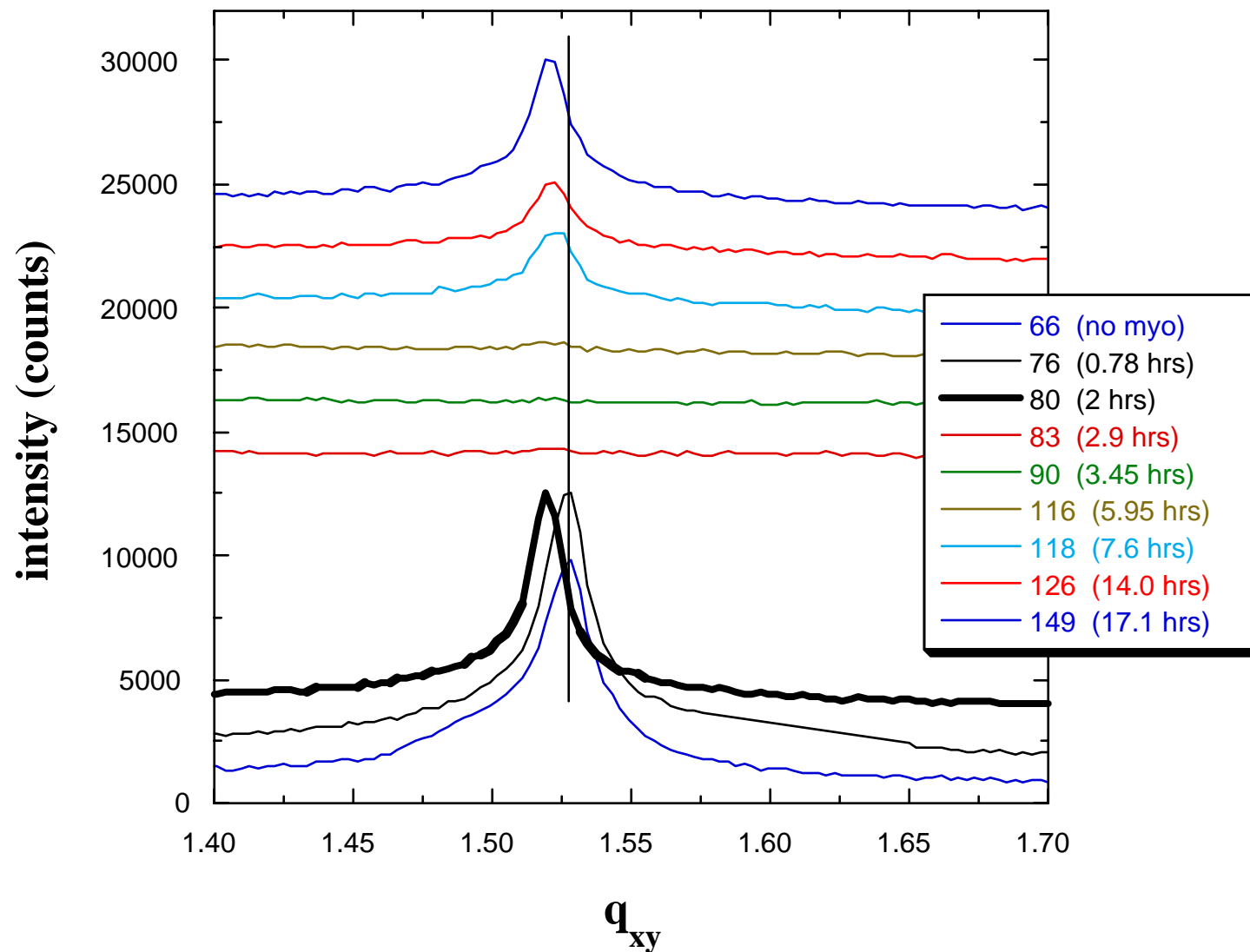
Constant pressure

3.7 hrs after injecting myoglobin

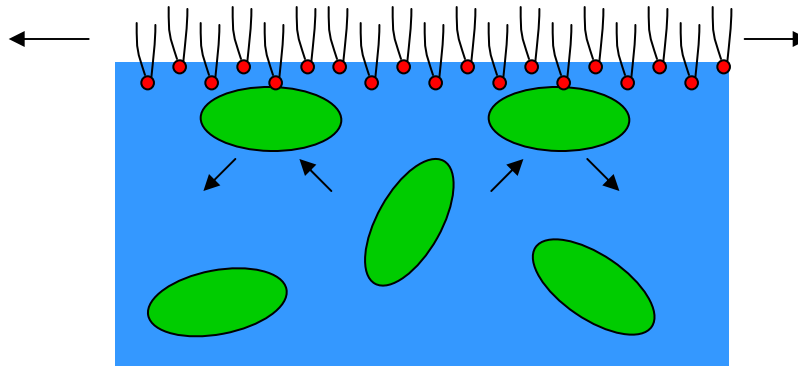


protein layer can be observed! thickness = 40 \AA , vol. fract. = 0.31

Ni²⁺, 40 mN/m - Bragg Peak

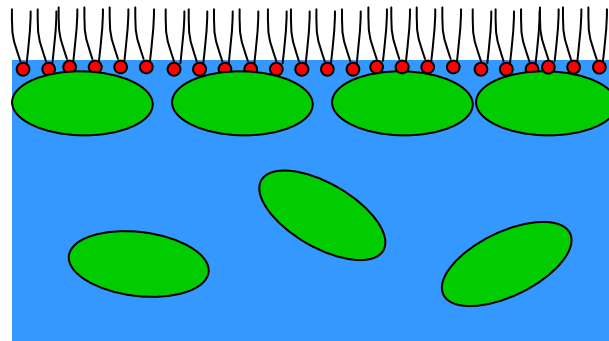


Proposed interpretation



(a)

Initial stage - reversible
adsorption



(b)

2nd stage - irreversible
adsorption and lipid
recrystallization



Summary

Protein associations with lipid membranes:

Grazing incidence scattering techniques provide insight into:

- evolution of adsorbed layer structure

- protein orientation

- denaturation upon adsorption

- effect of protein interactions on lipid film structure

 - initial stage - reversible**

 - later stage - irreversible**



Acknowledgements

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APS - CMC CAT
NIST - NG7
LANSCE - SPEAR